

*Week 10*

# ***Image Data Analysis(2)***



Today

Open images with matlab

Reading image and video files

Thresholding

Convolution

Segmentation

1. Thresholding

2. Watershed

# How to open image files?

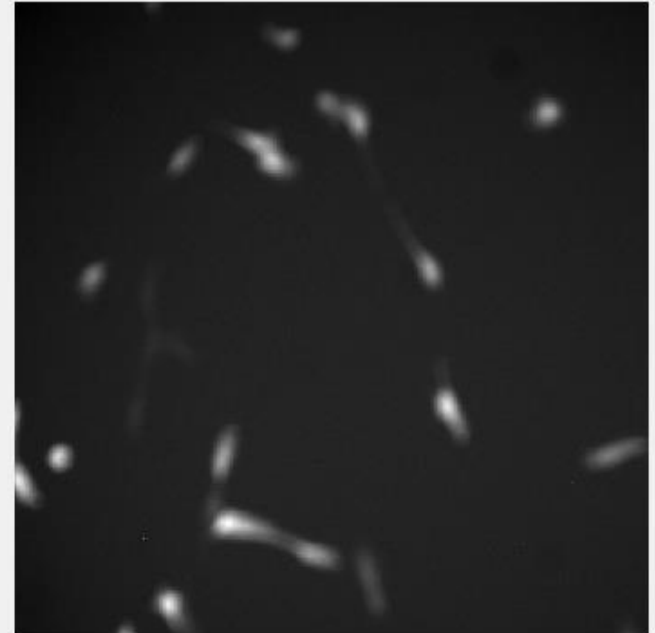
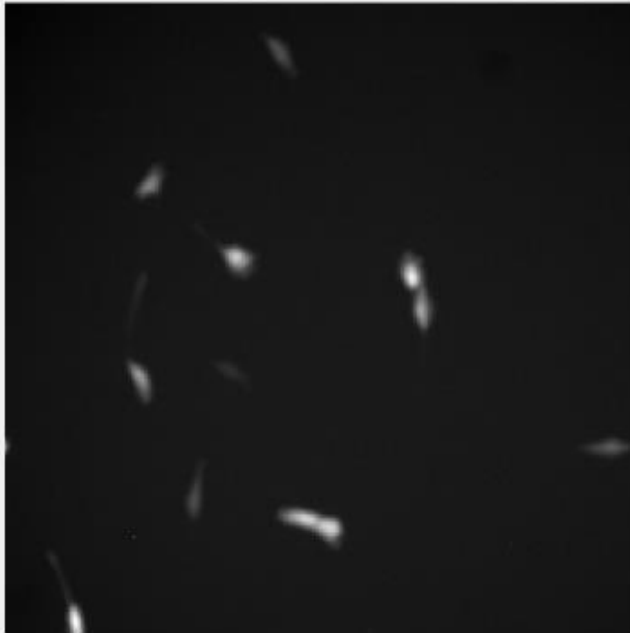
Functions: dir, imfinfo

```
cd 'C:\Users\90539\Documents\Teaching\MBG5X\Exampledatasets_nurhan_KODMSON_new'  
  
Spfile=dir('*.*tif')  
moivepro = imfinfo(Spfile(5).name);  
  
xsize = moivepro(1).Width;  
ysize = moivepro(1).Height;  
imageData = zeros(ysize, xsize, 1);  
%%
```

# A single file have many image frames

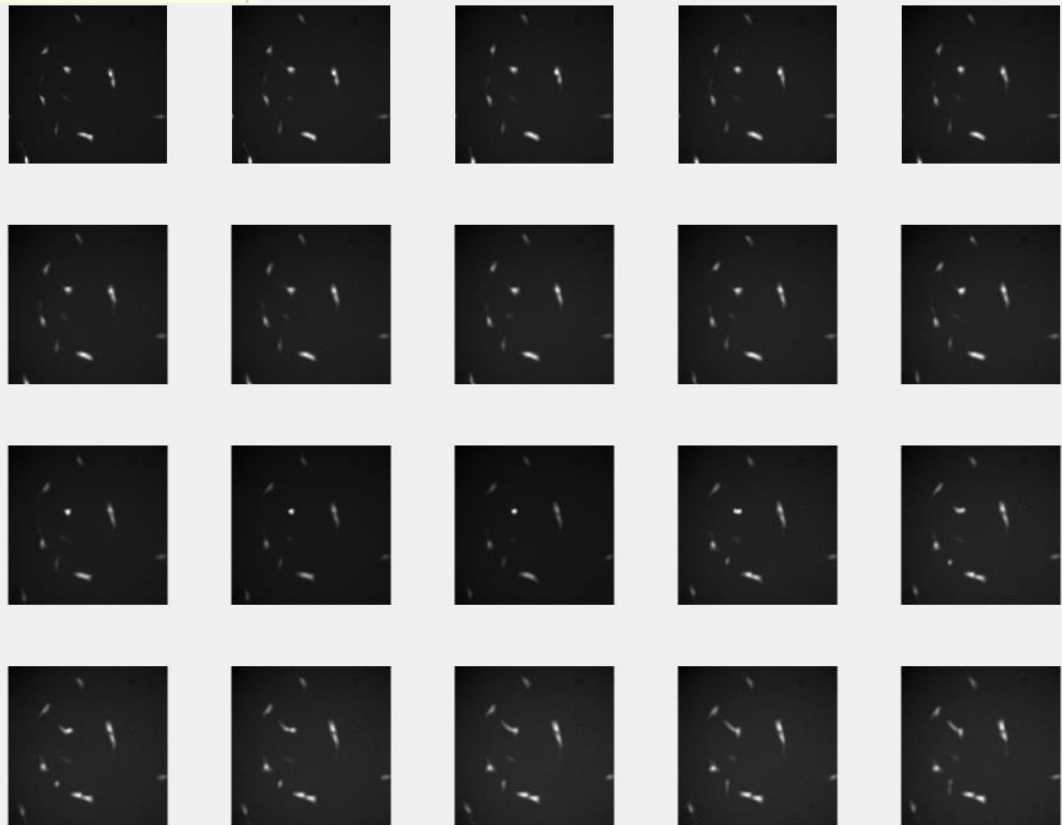
Functions: imread

```
imageData = zeros(512, 512, 1);  
%%  
for i=1:100;  
    fname= flistmovie{1};  
    imageData(:, :, i) = imread(fname, 'tif', i);  
end  
%%  
figure(1)  
subplot(1,2,1)  
imshow(imageData(:, :, 1), [])  
subplot(1,2,2)  
imshow(imageData(:, :, 80), [])  
%%
```



26  
27  
28  
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33  
34  
35  
36

```
%%  
figure(3)  
for i=1:20;  
    subplot(4,5,i)  
    imshow(imageData(:, :, i), [])  
    hold on  
end
```



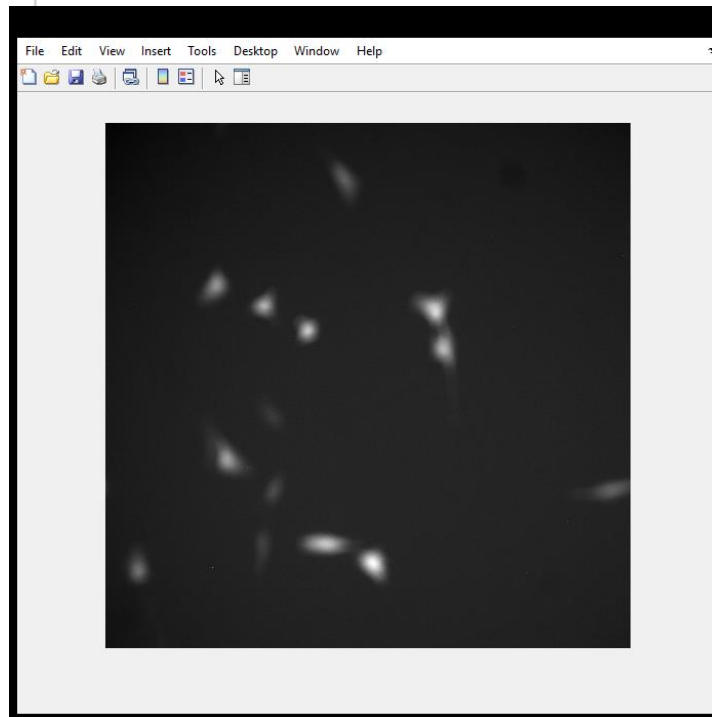
# Insert the image file into cell array

```
%%  
  
imgLD{1}=double(imageData(:,:,1:80))  
smin=min(min(imgLD{1}(:,:,1)))  
smax=max(max(imgLD{1}(:,:,1)))  
sratio=smax/smin  
[sr,sc,s]=find(imgLD{1}(:,:,1)==smax)
```

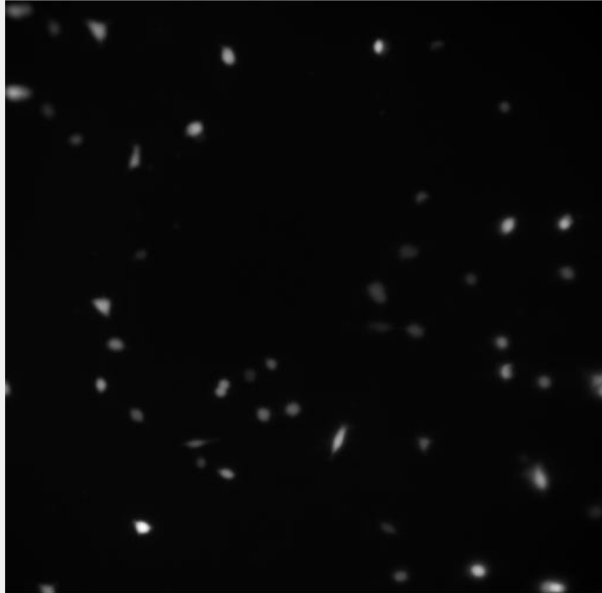
```
%%
```

Now run the movie file with the following code

```
%%  
figure(1)  
for i=1:80  
  
    imshow(imgLD{1}(:,:,1:end,i),[])  
    hold on  
    pause(0.2)  
end
```



# Open hela cell movies



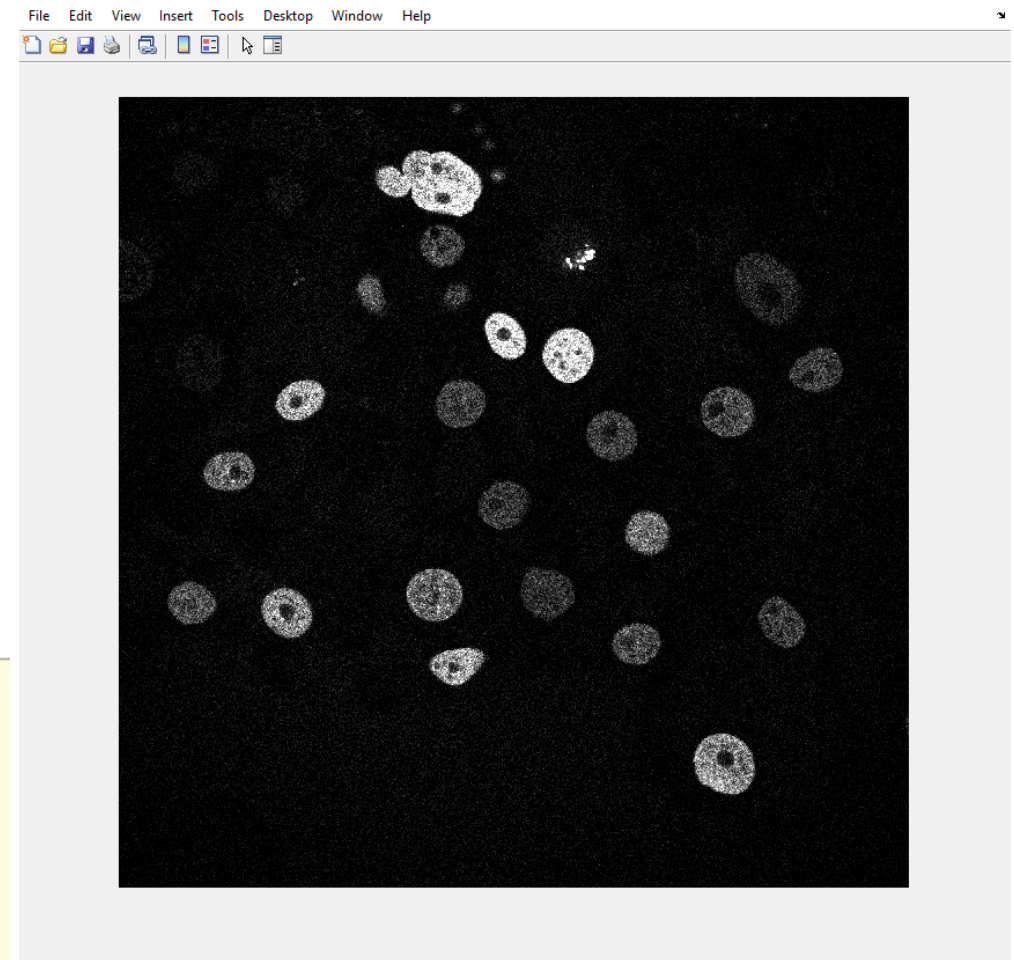
```
Lecture_4_openimages_videos.m
56
57 % loadinf other movies
58 Spfile=dir('032 GFP.tif')
59 flistmovie = {Spfile(1).name};
60
61 moivepro = imfinfo(flistmovie{1});
62 xsize = moivepro(1).Width;
63 ysize = moivepro(1).Height;
64 imageData = zeros(ysize, xsize, 1);
65 imageData = imread( flistmovie{1}, 'tif', 1);
66 for i=1:40;
67
68     fname= flistmovie{1};
69     imageData(:, :, i) = imread(fname, 'tif', i);
70 end
71 %imgLD{1}=double(imageData)
72 imgLD{1}=double(imageData(:, :, 1:40))
73 smin=min(min(imgLD{1}(:, :, 1)))
74 smax=max(max(imgLD{1}(:, :, 1)))
75 sration=smax/smin
76 [sr,sc,sl]=find(imgLD{1}(:, :, 1)==smax)
77 %
78
79 for i=1:1
80 figure(1)
81 imshow(imgLD{1}(:, 1:end, 1),[])
82 hold on
83 pause(0.5)
84 end
85
86 %%%
```

# Sometimes movies files are saved as a individual image files

Current Folder

Name	Size	Date Modified	Type
t000.tif	267 KB	11/23/2012 2:16 PM	TIF File
t002.tif	265 KB	11/23/2012 2:16 PM	TIF File
t003.tif	261 KB	11/23/2012 2:16 PM	TIF File
t004.tif	259 KB	11/23/2012 2:16 PM	TIF File
t005.tif	260 KB	11/23/2012 2:16 PM	TIF File
t006.tif	266 KB	11/23/2012 2:16 PM	TIF File
t007.tif	265 KB	11/23/2012 2:16 PM	TIF File
t008.tif	264 KB	11/23/2012 2:16 PM	TIF File
t009.tif	260 KB	11/23/2012 2:16 PM	TIF File
t010.tif	256 KB	11/23/2012 2:16 PM	TIF File
t011.tif	255 KB	11/23/2012 2:16 PM	TIF File
t012.tif	263 KB	11/23/2012 2:16 PM	TIF File
t013.tif	262 KB	11/23/2012 2:16 PM	TIF File
t014.tif	261 KB	11/23/2012 2:16 PM	TIF File
t015.tif	260 KB	11/23/2012 2:16 PM	TIF File
t016.tif	253 KB	11/23/2012 2:16 PM	TIF File
t017.tif	251 KB	11/23/2012 2:16 PM	TIF File
t018.tif	259 KB	11/23/2012 2:16 PM	TIF File

Details



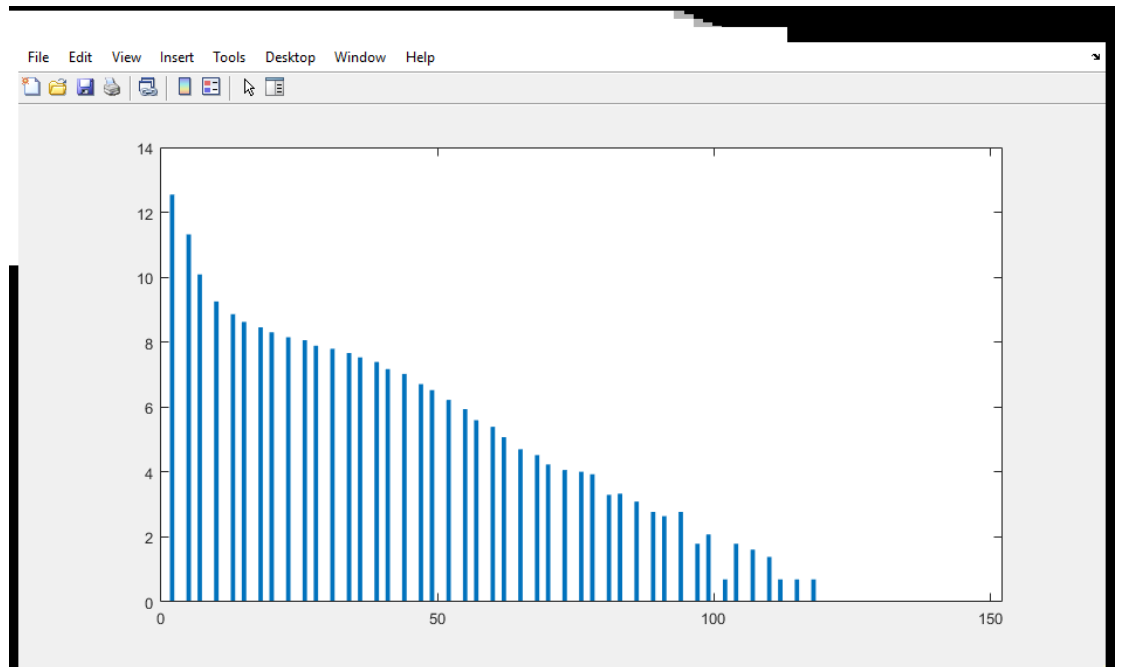
```
%%  
for i=1:nframes;  
    % read the name  
    imageData(:,:,i) = imread(Spfile(i).name, 'tif', 1);  
end  
  
imgLD{1}=double(imageData);  
  
figure(1)  
imshow(imgLD{1,1}(:,:,40),[min(min(imgLD{1,1}(:,:,1))) max(max(imgLD{1,1}(:,:,1)))/4.0]  
max(max(imgLD{1,1}(:,:,1)))  
min(min(imgLD{1,1}(:,:,1)))  
saveas(gcf,'originalfiguresim_011.tif')
```



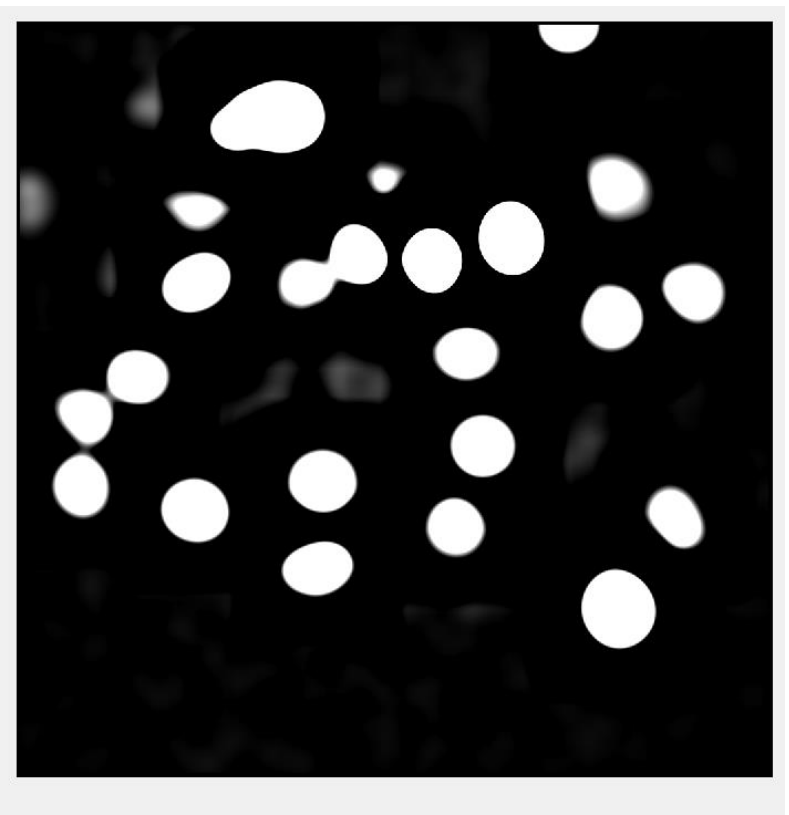
# Image histograms

Functions: reshape, find, histcounts

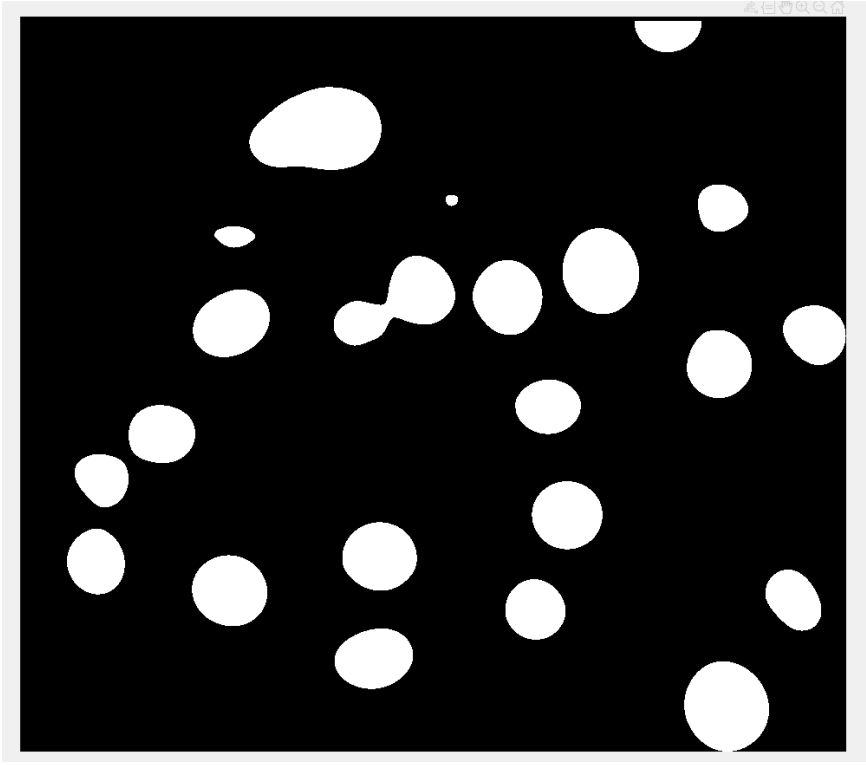
```
%%  
x= reshape(imgLD{1}(:,:,1),1,[]);  
y=sort(x,'ascend')  
  
[r,c,]=find(y>0)  
  
edges=1:1:Lcutmax  
n=histcounts(y(1,c(1,1):end),edges)  
  
figure(2)  
bar(edges(1:151), log(n))  
%%
```



# Watershed based segmentation



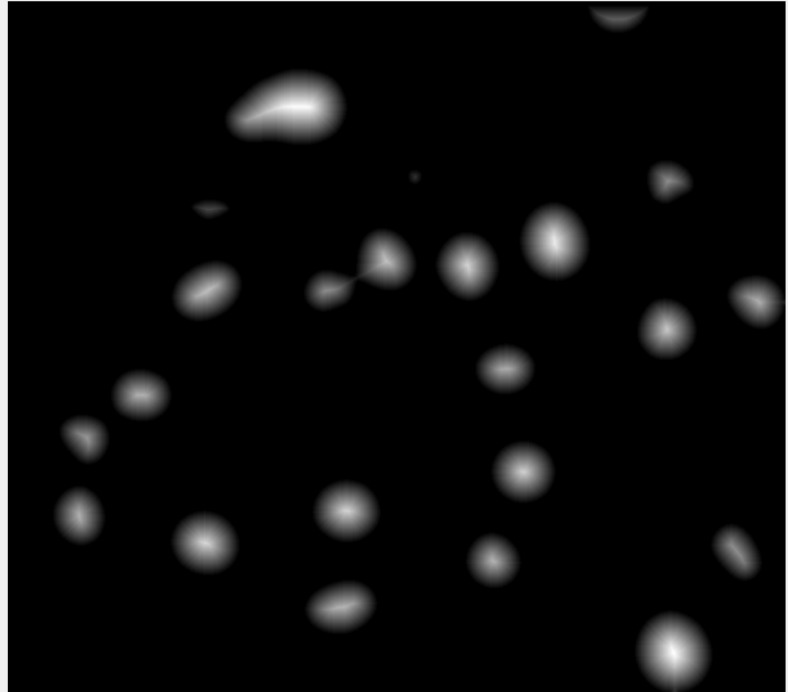
After convolution



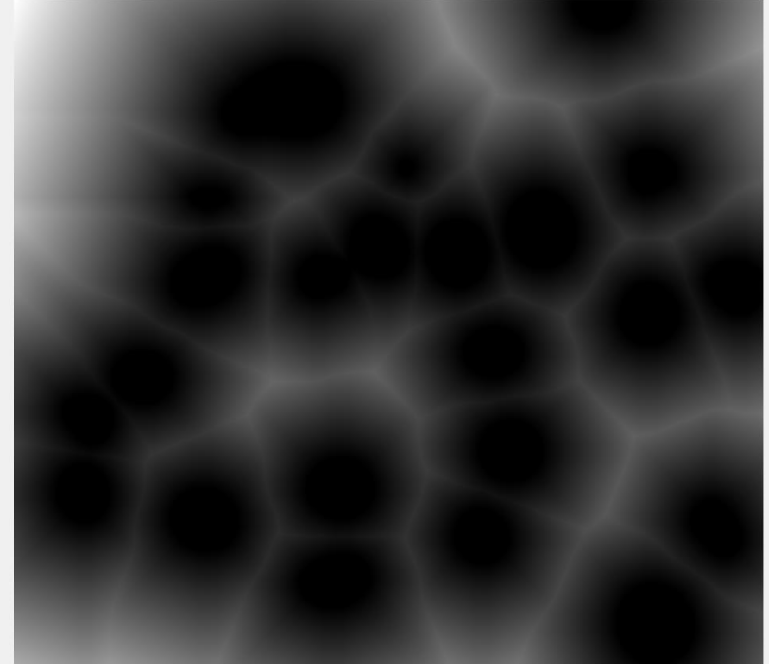
Binary image after thresholding

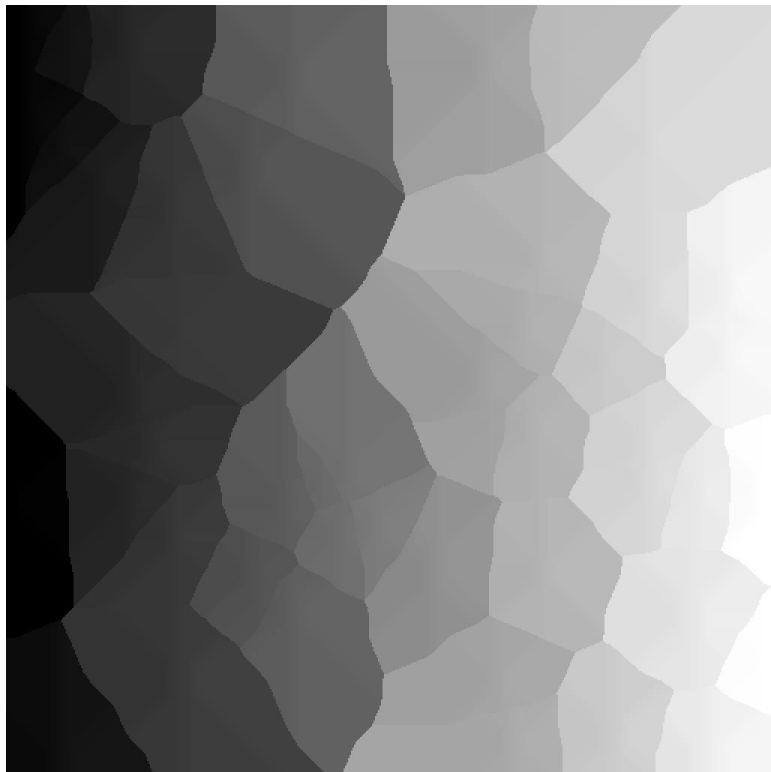
# Bwdist, used for the distance transform of image, Distance between pixel and nearest nonzero pixel

```
%%  
bw=imgetes;  
  
D = bwdist(~imgetes,'quasi-euclidean');  
%D = bwdist(~imgetes,'chessboard');  
%figure(24)  
%imshow(D,[])  
%title('Distance Transform of Binary Image')  
D = D;  
figure(25)  
imshow(D,[])  
0% 0%
```



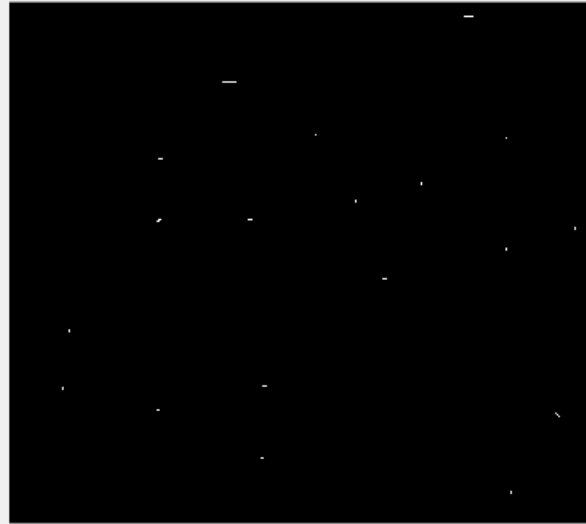
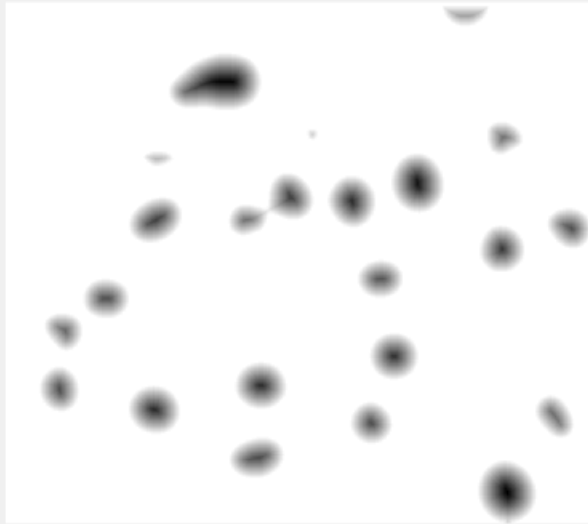
```
%%  
bw=imgetes;  
  
D = bwdist(imgetes,'quasi-euclidean');  
%D = bwdist(~imgetes,'chessboard');  
%figure(24)  
%imshow(D,[])  
%title('Distance Transform of Binary Image')  
D = D;  
figure(25)  
imshow(D,[])  
0% 0%
```





```
%%  
bw=imgetes;  
  
[D,idx] = bwdist(imgetes,'quasi-euclidean');  
%D = bwdist(~imgetes,'chessboard');  
%figure(24)  
%imshow(D,[])  
%title('Distance Transform of Binary Image')  
Dx = -D;  
figure(25)  
subplot(1,2,1)  
imshow(Dx,[])  
subplot(1,2,2)  
imshow(D,[])  
  
figure(26)  
imshow(idx,[])
```

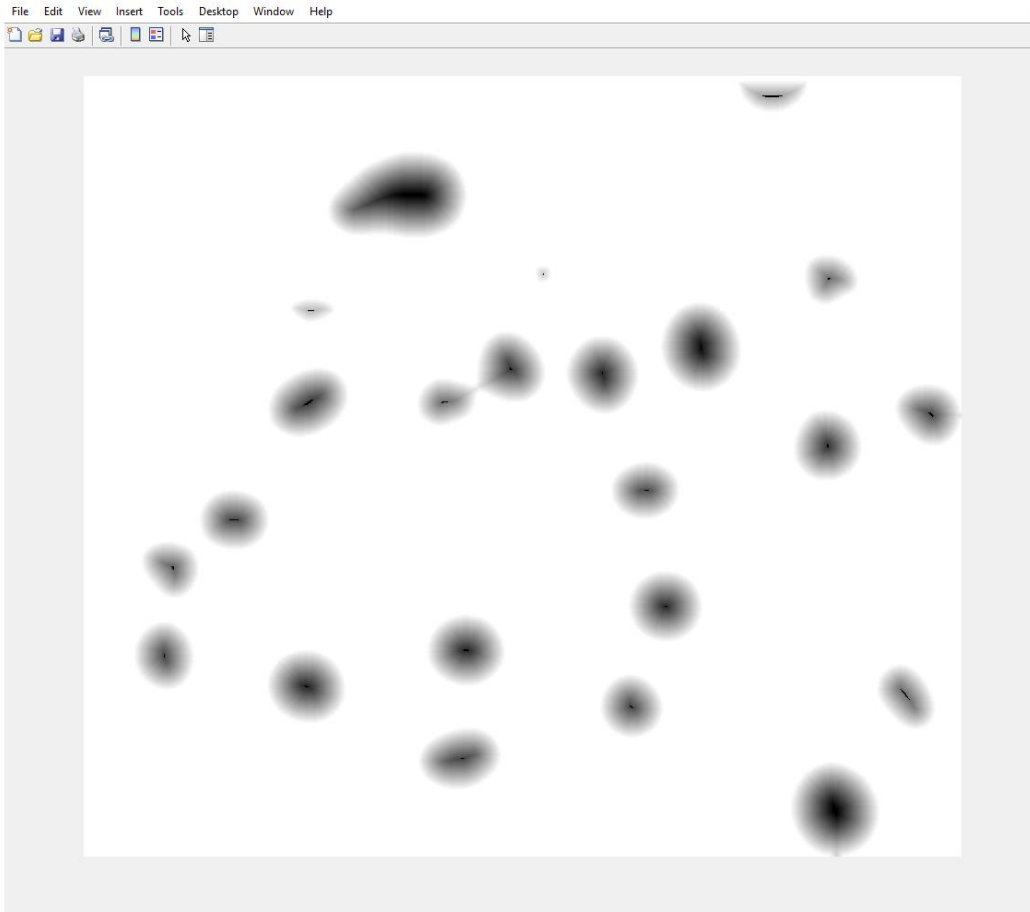
Here we compute the regional minima



```
%%  
%title('Complement of Distance Transform')  
%  
%step 2  
mask = imextendedmin( (-1*min(min(D))) * D, 36);  
% figure(34)  
% imshowpair(bw,mask,'blend')  
%  
figure(35)  
subplot(1,2,1)  
imshow(D,[])  
subplot(1,2,2)  
imshow(mask,[])
```

Here we find the global minimum for each region

# Computation of marked image



---

```
%%
```

```
D2 = imimposemin(D,mask);
```

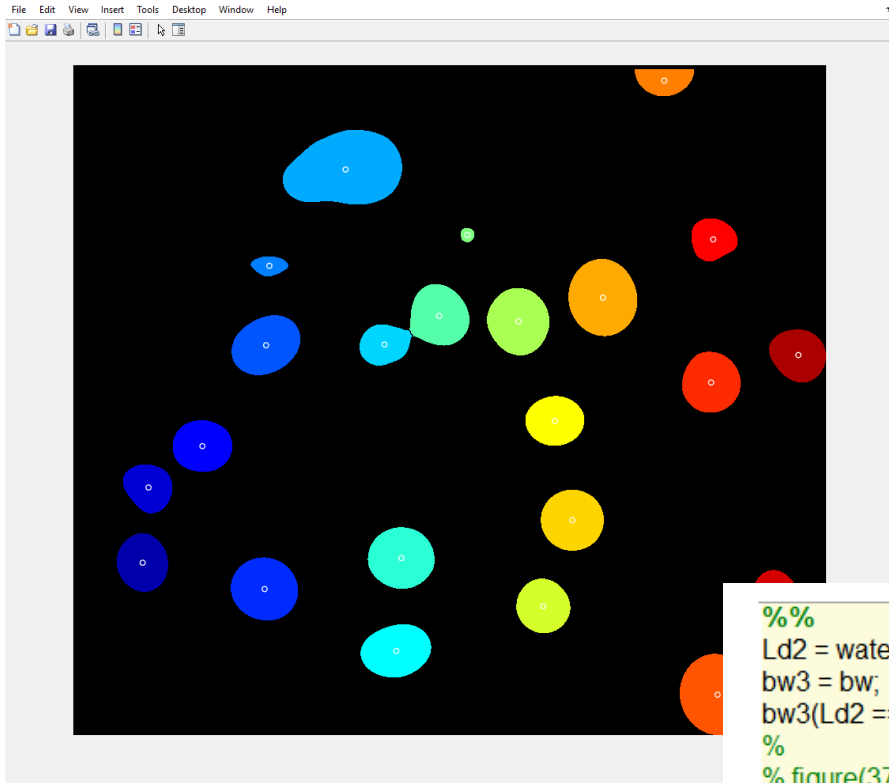
```
figure(36)
```

```
imshow(D2,[])
```

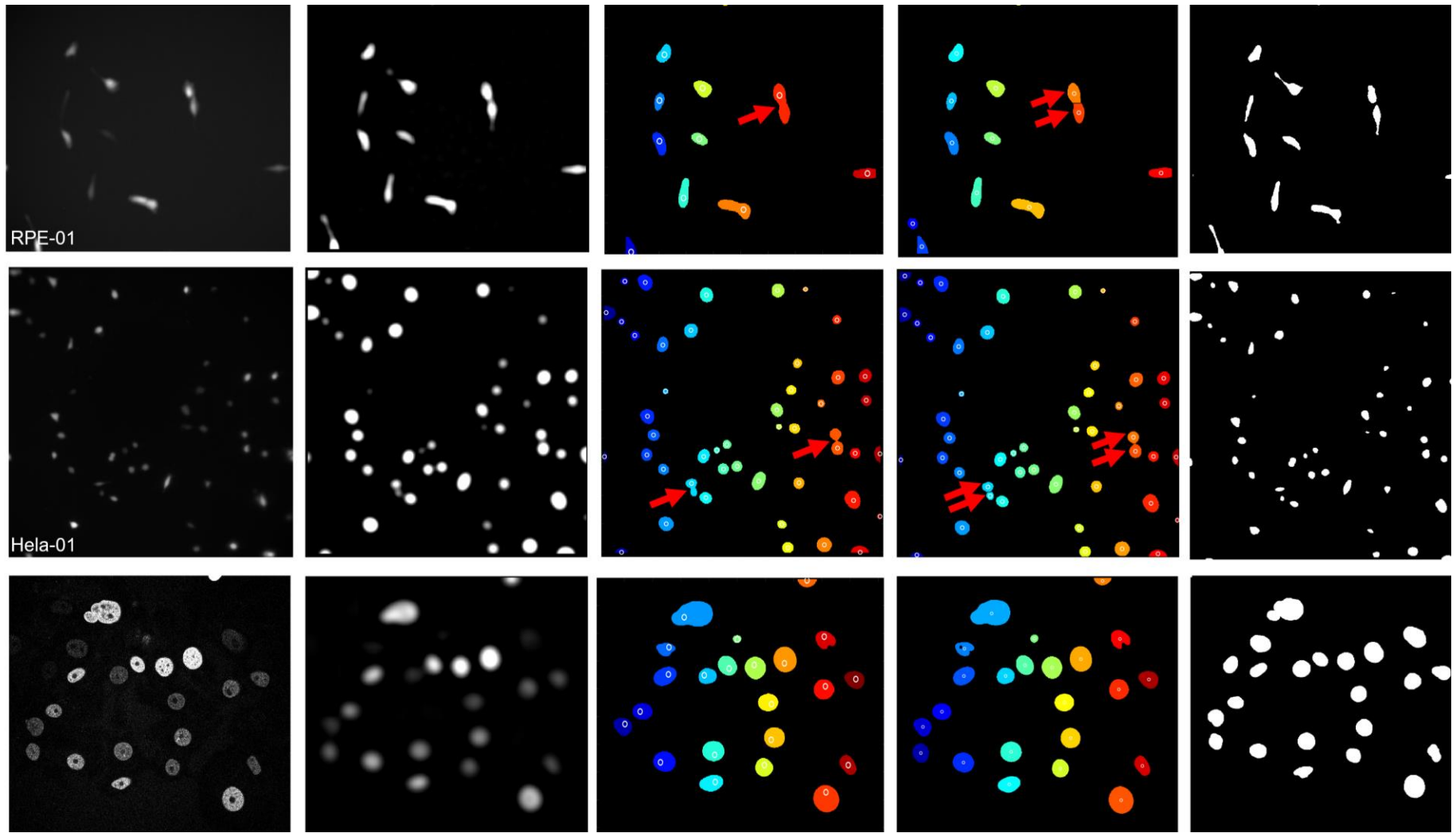
---

```
end
```

# Here is the watershed image



```
%%  
Ld2 = watershed(D2);  
bw3 = bw;  
bw3(Ld2 == 0) = 0;  
%  
% figure(37)  
% subplot(1,2,1)  
% imshow(bw3,[])  
% subplot(1,2,2)  
% imshow(Ld2,[])  
  
label1 = bwlabel(bw3);  
rgb = label2rgb(label1,'jet',[0 0 0]);  
  
stats = regionprops("table",label1,"Centroid", "MajorAxisLength","MinorAxisLength")  
figure(3)  
imshow(rgb)  
hold on  
plot(stats.Centroid(:,1), stats.Centroid(:,2),'ow','linewidth',1,'MarkerSize',5)
```

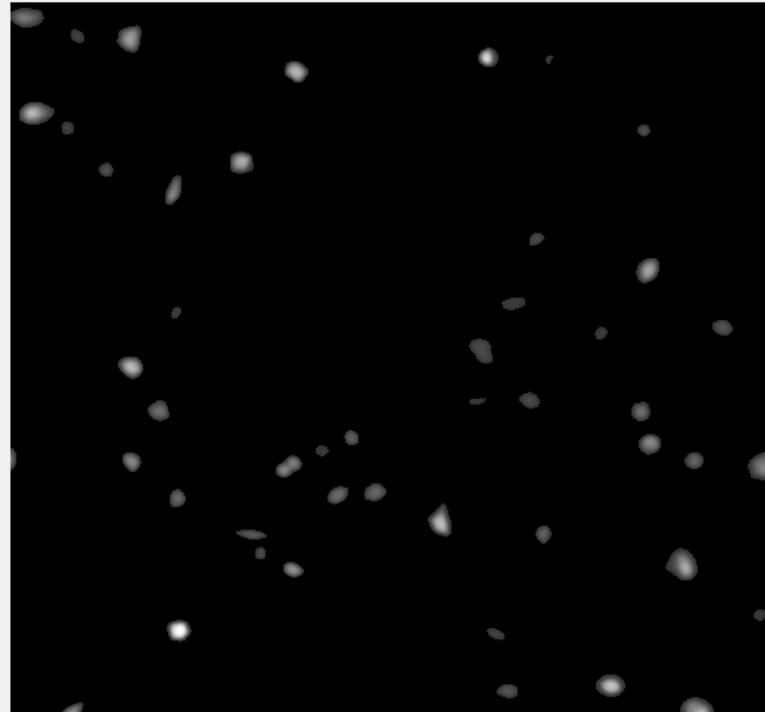
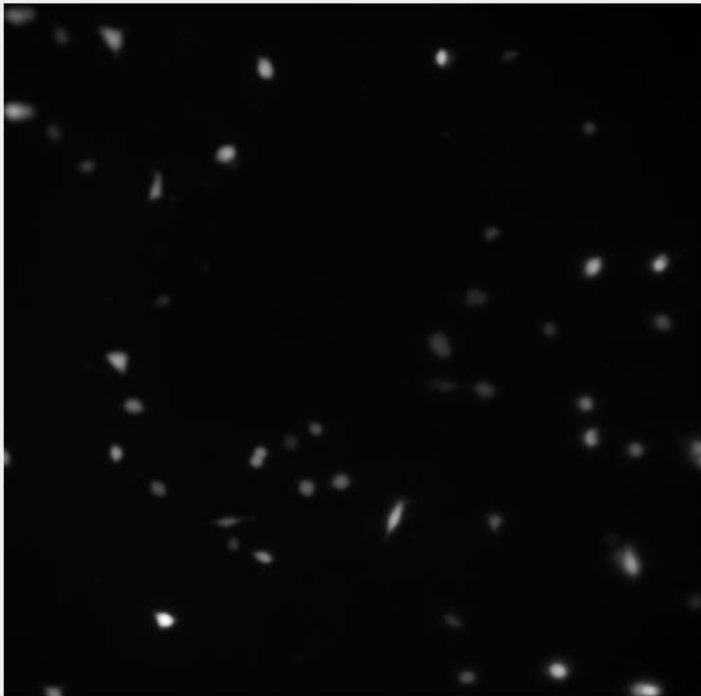




# Analysis of Hela Cells

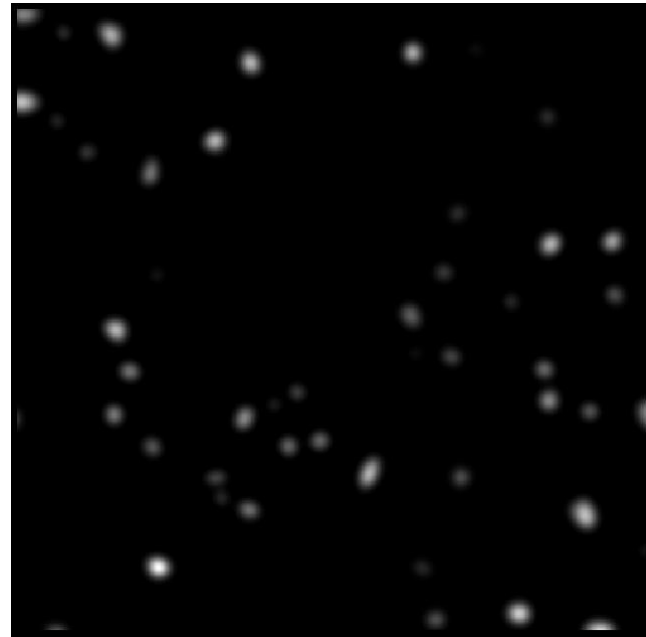
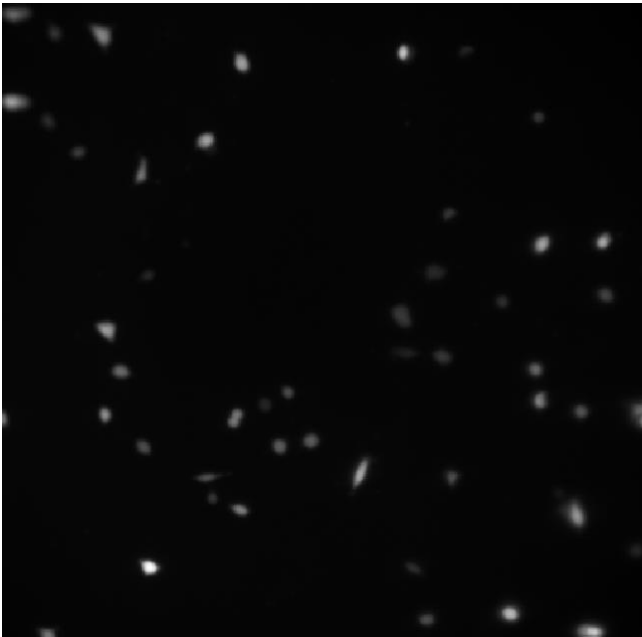
```
for i=1:nframes;  
    Lcut=min(min(imgLDout{i,1}));  
    background=Lcut*2  
    imgLDout{i,1}(imgLDout{i,1}<2000)=0;  
end  
figure(9)  
imshow(imgLDout{3,1},[])
```

File Edit View Insert Tools Desktop Window Help



# Convolved image

```
360
361 %%
362 clear imageOut3
363 objectSize=50;
364 noiseSize=3
365 for i=1:nframes;
366
367     imageIn = imgLDout{i,1};
368     [imageOut3{i,1}]=objFilter5son(imageIn,objectSize, noiseSize);
369 end
370
371 for i=1:1
372     figure(100)
373     subplot(1,2,1)
374     imshow(imgLD{1,1}{:,i},[])
375     subplot(1,2,2)
376     imshow(imageOut3{i,1}{:,},[])
377
378 end
379
```



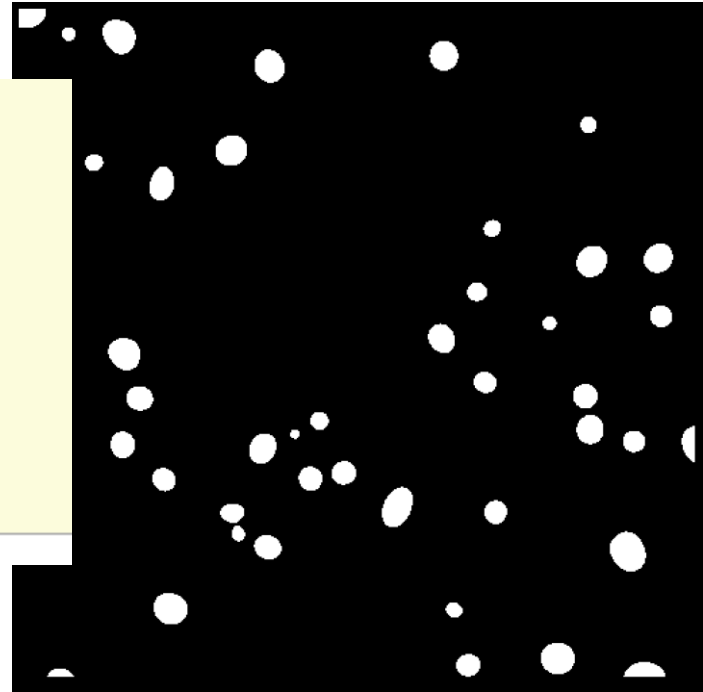
# conv2

It is used to remove pixel noise and create circular objects

```
1 function [imageOut]=objFilter(imagelIn,objectSize,noiseSize)
2
3 % Software name: objFilter
4 %
5 % Software summary: To remove background pixel noise in time-lapse video frames
6 % using bandpass filter
7 %
8 % Brief description: This function can be used to apply bandpass filter for
9 % each image in video frames. The smoothed images are generated after remove pixel noise
10 % in images. To remove pixel noise, a gauss filter is accordingly computed
11 % by using noise size. Then it was applied in both dimensions.
12 %
13 % How to run code?
14 % To use this function, type the following line into Editor,
15 % Enter Sfile that includes the name of individual frames in a movie file,
16 % then frame number is needed and click Run.
17 %
18 % for i=1:nframes;
19 %
20 % imageIn = imageLoad1{1}{:,i};
21 % [imageOut{i,1}]=objFilter(imageIn,objectSize, noiseSize);
22 %
23 % end
24 % Inputs:
25 %
26 % 1. imageIn: Background subtracted 2D image frames
27 % 2. objectSize: An average length of objects in frames. The default value is 50 pixel (10x obje
28 % User has to open a single image and measure the length of a few cells in
29 % pixel. Enter an average cell size.
30 % 3. noiseSize: Average length of noise. Set the pixel value as an integer number. The default
31 % User has to check a few frames and determine the average diameter of
```

Now we can segments the image by finding the connected pixels

```
380 %%  
381 % check image before separate pixels  
382 - katdegeri=1.10  
383 - rang=katdegeri*std(imageOut3{1,1}(:));  
384 - [rPixel,cPixel,lo]=find(imageOut3{1,1}(:,.) > rang);  
385 - imagetes=zeros(1024,1024);  
386 - imagetes=zeros(512,512);  
387 - for i=1:size(rPixel,1);  
388 -     imagetes(rPixel(i,1),cPixel(i,1))=1;  
389 - end  
390 - figure(13)  
391 - imshow(imagetes,[])  
392 %%
```



```
393 %%  
394 % here compute the cell regions  
395 % segmentation step based on colvoluted image  
396 - tic  
397 - clear output1  
398 - for i=1:1;  
399 -     i  
400 -     [output1{ 1,1},output1{1,2}]=objExplorer2son(imageOut3{i,1},katdegeri,i); %028  
401 -  
402 - end  
403 - toc
```

```

404 %%
405 l=size(imageOut3{1,1},1)
406 m=size(imageOut3{1,1},2)
407 imagetest=zeros(l,m);
408 cellidx=output1;
409 k=1
410 for ts=1:size(cellidx{1,1},1);
411     % here we fill the array with cell numbers from 1 to n
412     % we use the coordinates data from
413     for i=1:size(cellidx{1,2},1);
414         if cellidx{1,2}(i,3)==ts;
415             imagetest(cellidx{1,2}(i,1),cellidx{1,2}(i,2))=k;
416             end
417         end
418         k=k+1
419     end
420 end
421
422 objSgmt{1,1}=imagetest;
423 colMatList=[0,0,0];
424 colMatList(2:257,1:3)=jet(256);
425
426 figure(2)
427 imagesc(imagetest)
428 colormap(colMatList)
429 colorbar
430 hold on
431 plot(cellidx{1,1}(1:end,1), cellidx{1,1}(1:end,2),'ow','linewidth',1,'Markersize',5)
432 saveas(gcf,'sim1segmentedimage2.tif')
433 save('segmenteddatasim2','output1')
434
435

```

